

SYNTHESES OF (α 2-9) AND (α 2-8) LINKED NEURAMINYLNURAMINIC ACID
DERIVATIVES¹

Kaoru Okamoto,[†] Tadao Kondo,^{††*} and Toshio Goto^{*}

Laboratory of Organic Chemistry, Faculty of Agriculture; and

^{††}Chemical Instrument Center; Nagoya University

Chikusa, Nagoya 464, Japan

Abstract: A newly prepared glycosyl donor, the acetyl protected 2 β -bromo-3 β -hydroxy-N-acetylneuraminic ester was condensed with the protected 2-deoxy-2,3-dehydroneuraminic ester having a free hydroxy group at 9- or 8-position to form NeuAc(α 2-9)NeuAc and NeuAc(α 2-8)-NeuAc linkage, respectively, which are involved in the group C meningococcal polysaccharides and gangliosides. The obtained 3 β -hydroxy disaccharides were phenoxythiocarbonylated, reduced with tri-*n*-butylstannane, and deprotected to give the free glycosides in high yields.

N-Acetylneuraminic acid (NeuAc) located at the non-reducing ends of carbohydrate chains of glycoproteins and glycolipids plays a role in biological recognition.^{2,3} Many attempts have been done with little success in the synthesis of NeuAc linked carbohydrate chains. In the previous paper,⁴ we have shown that the 2 β -bromo-3 β -hydroxy-NeuAc methyl ester **1**⁵ was a useful glycosyl donor to afford NeuAc(α 2-3)Gal or NeuAc(α 2-3')Lac glycosides. We report here a successful application of this approach to the syntheses of NeuAc(α 2-9)NeuAc and NeuAc(α 2-8)NeuAc linkages involved in the group C meningococcal polysaccharides⁶ and gangliosides. This is the first synthesis of the NeuAc(α 2-8)NeuAc linkage.

The glycosylation of the 9-unprotected 2,3-dehydro-NeuAc methyl ester **4**⁷ (1.1 equiv) with the 2-bromo-3-hydroxy derivative of peracetylated NeuAc methyl ester **1** in toluene-1,2-dichloroethane (1:1) in the presence of silver triflate (AgOTf) at 0 °C gave in 71% isolated yield a mixture of 2-9 linked di-NeuAc derivatives. The mixture was separated by the repeated silica gel column chromatography (benzene-acetone-methanol, gradient elution from 30:30:0 to 30:30:2) or by preparative ODS HPLC (methanol-water, 46:54) to the protected NeuAc-

[†]Present address: Institute of Bio-Active Science (IBAS), Nippon Zoki Pharmaceutical Co., Ltd., Kinashi, Yashiro-cho, Kato-gun, Hyogo 673-14, Japan.

(α -9)NeuAc glycoside **6**⁹ (mp 137-139 °C, 42% yield) and the corresponding β -glycoside **10**⁹ (21% yield). As demonstrated earlier, the anomeric configuration was determined by analysis of ¹H-NMR spectrum. Thus, the $J_{7,8}$ coupling constant of NeuAc units of **6** and **10** showed 8.4 and 2.7Hz, respectively, and $\Delta\delta|H-9'-H-9|$ values of them were 0.17 and 0.59ppm, respectively. These values agreed with those deduced from the empirical rule.¹⁰

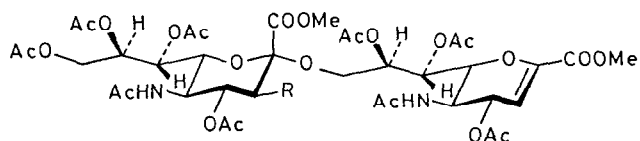
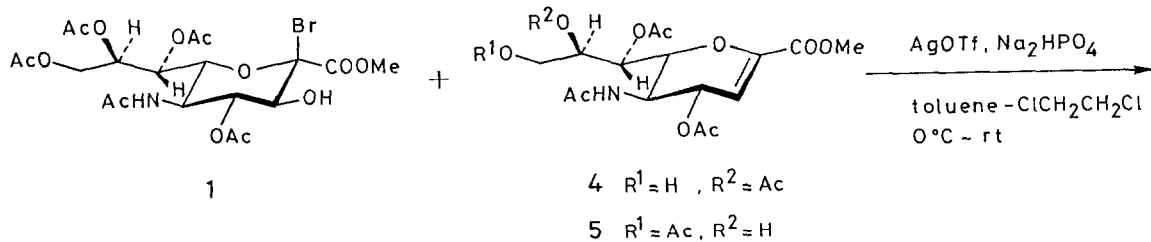
In the above glycosylation condition, the bromide **1** reacted with the 8-unprotected 2,3-dehydro-NeuAc methyl ester **5**¹¹ (1.0 equiv) giving a mixture of 2-8 linked di-NeuAc derivatives in 43% yield. The mixture was separated by preparative ODS HPLC (methanol-water, 45:55) to the NeuAc(α 2-8)NeuAc glycoside **14**⁹ (26% yield) and the NeuAc(β 2-8)NeuAc glycoside **18**⁹ (8% yield). The anomeric configuration was determined by the fact that in ¹H-NMR spectra the $J_{7,8}$ coupling constant of **14** and **18** showed 7.5 and 1.8Hz, respectively, and $\Delta\delta|H-9'-H-9|$ values of them were 0.41 and 0.91ppm, respectively.

The 3 β -hydroxy groups of the glycosides **6**, **10**, **14**, and **18** were reduced to the corresponding 3-deoxy glycosides in the following way. The glycosides were treated with phenyl chlorothionocarbonate and 4-dimethylaminopyridine¹² in acetonitrile for **6** and **10**, in DMF for **14**, and in DMSO for **18** to give the thionocarbonates **7**⁹ (91% yield, $J_{7,8}$ =8.9Hz, $\Delta\delta|H-9'-H-9|$ =0.06ppm), **11**⁹ (83%, 4.0Hz, 0.62ppm), **15**⁹ (85%, 7.5Hz, 0.42ppm), and **19**⁹ (80%, 2.7Hz, 0.85ppm), respectively. Reduction of the thionocarbonates with tri-*n*-butylstannane afforded **8**⁹ (97% yield, $J_{7,8}$ =8.9Hz, $\Delta\delta|H-9'-H-9|$ =0.18ppm), **12**⁹ (96%, 3.4Hz, 0.67ppm), **16**⁹ (95%, 7.8Hz, 0.53ppm), and **20**⁹ (96%, 2.7Hz, 0.91ppm), in which **20** was identical with the previously obtained authentic sample.¹¹ The $J_{7,8}$ coupling constant and $\Delta\delta|H-9'-H-9|$ value remained unaltered in each series of α - and β -glycosides, even if the C-3 was substituted with hydroxy or phenoxythiocarbonyl group. Deacetylation (*t*-BuOK in methanol at room temp) and saponification (1N NaOH in methanol at room temp) of **8**, **12**, and **16** afforded the deprotected di-NeuAc **9**,⁹ **13**,⁹ and **17**,⁹ quantitatively.

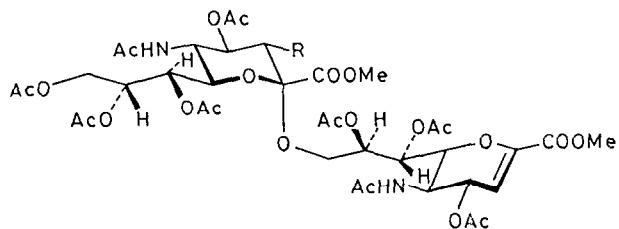
In conclusion, we found that the 2 β -bromo-3 β -hydroxy-NeuAc derivative **1** is a useful donor for the glycosylation of the 8- or 9-unprotected peracetyl 2,3-dehydro-NeuAc methyl ester since the 3 β -hydroxy group of the bromide **1** prevents the dehydrohalogenation reaction and assists the glycosylation reaction.¹³ The 2,3-double bond present in the glycosylation products can be used for further glycosylation via the corresponding 2 β ,3 β -bromohydrin.⁵

REFERENCES AND FOOTNOTES

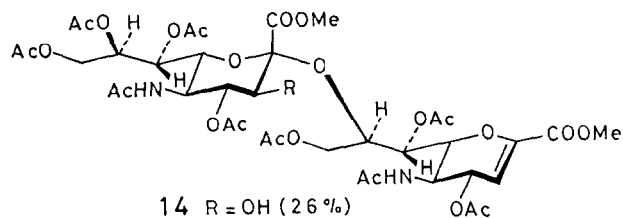
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6 $R = \text{OH}$ (42%)7 $R = \text{OC}(=\text{S})\text{Ph}$ (91%)8 $R = \text{H}$ (97%)

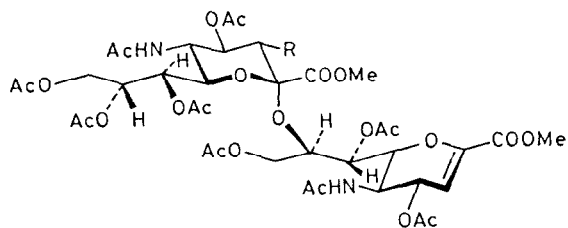
9 deprotected 8 (97%)

10 $R = \text{OH}$ (21%)11 $R = \text{OC}(=\text{S})\text{Ph}$ (83%)12 $R = \text{H}$ (96%)

13 deprotected 12 (96%)

14 $R = \text{OH}$ (26%)15 $R = \text{OC}(=\text{S})\text{Ph}$ (85%)16 $R = \text{H}$ (95%)

17 deprotected 16 (97%)

18 $R = \text{OH}$ (8%)19 $R = \text{OC}(=\text{S})\text{Ph}$ (80%)20 $R = \text{H}$ (96%)

this issue.

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7. The glycosyl acceptor **5** having a free hydroxy group at 9-position was prepared from 2-deoxy-2,3-dehydro-NeuAc methyl ester **2**⁸ in the following two steps: (i) trityl chloride in pyridine at 60 °C for 2 h, then Ac₂O-pyridine at 60 °C for 1 h (4,7,8-tri-O-acetyl-9-O-Tr **3**⁹, 84%); (ii) 90% AcOH at 60 °C for 1 h (9-ol **4**⁹, 71%).
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9. Satisfactory elemental analyses were obtained for these compounds. $[\alpha]_D$ and ¹H-NMR data (non-reducing NeuAc unit in chloroform-d) are shown below.

Com- pound	$[\alpha]_D^a$	Chemical shifts (δ) and J _{7,8} coupling constants, Hz in ¹ H-NMR											
		H-3eq (dd)	H-3ax (dd)	H-4 (dd)	H-5 (ddd)	H-6 (dd)	H-7 (dd)	H-8 (ddd)	H-9 (dd)	H-9' (dd)	Me ester (s)	NH (d)	J _{7,8}
3	+38.3°	6.05 ^b	5.50	4.27	4.48	5.58	5.37	3.24	3.40	3.75	5.66	5.5	
4	+58.7°	5.98 ^b	5.52	4.49	4.39	5.46	5.11	3.61 ^c	4.05	3.81	5.91	5.6	
6	+29.9°	3.77	5.22	4.16	4.62	5.27	5.36	4.08	4.25	3.81 ^d	5.73	8.4	
7	+35.3°	5.68 ^b	5.73	4.19	4.88	5.30	5.48	4.17	4.23	3.80 ^d	5.72	8.9	
8	+17.8°	2.53	1.89	4.94 ^c	3.97	4.13	5.33	5.40	4.12	4.30	3.80 ^d	5.37	8.9
9 ^e	+12.7° ^f	2.70	1.80	3.77 ^c	3.88 ^g	3.81	3.54	3.87	3.62	3.85		8.9	
10	+23.9°	3.81	5.24	4.16	4.29	5.37	5.05	4.08	4.67	3.82 ^d		2.7	
11	+35.4°	5.82 ^b	5.54	4.38	4.44	5.42	5.19	4.09	4.71	3.80 ^d	6.52	4.0	
12	+35.1°	2.44	1.84	5.28 ^c	4.05	4.30	5.41	5.17	4.13	4.80	3.81 ^d	6.26	3.4
13 ^e	+6.6° ^f	2.45	1.76	4.10 ^c	3.93 ^g	3.93	3.56	3.84	3.64	3.83		9.5	
14	+27.4°	4.00	5.24	4.19	4.38	5.24	5.28	3.96	4.37	3.83 ^d	5.67	7.5	
15	+35.4°	5.89 ^b	5.47	4.40	4.38	5.32	5.34	4.02	4.44	3.81 ^d	5.48	7.5	
16	+22.3°	2.65	2.19	4.88 ^c	4.00	3.81	5.31	5.31	3.95	4.48	3.83 ^d	5.14	7.8
17 ^e	+35.7° ^f	2.71	1.78	3.66 ^c	3.84 ^g	3.67	3.54	3.78	3.62	3.83		9.0	
18	+20.3°	3.85	4.99	4.21	4.66	5.35	5.32	3.95	4.86	3.82 ^d	6.03	1.8	
19	+22.9°	5.84 ^b	5.26	4.38	4.72	5.38	5.33	4.00	4.85	3.79 ^d	6.20	2.7	
20	+31.2°	2.49	1.84	5.09 ^c	4.08	4.62	5.37	5.31	4.01	4.92	3.78 ^d	6.06	2.7

^a Measured in chloroform. ^b Multiplicity: d. ^c Multiplicity: ddd.

^d Assignments may be interchanged with reducing unit. ^e Measured in D₂O (t-BuOH=1.23 ppm). ^f Measured in water. ^g Multiplicity: dd.

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13. When the 3-O-acetylated compound of **1** was used in this glycosylation, no glycosides was obtained.

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